



Europäisches Patentamt

European Patent Office

Office européen des brevets



11) Publication number: 0 580 362 A1

(12)

### **EUROPEAN PATENT APPLICATION**

(21) Application number: 93305569.1

(22) Date of filing: 15.07.93

(51) Int. Cl.5: G01N 21/64

(30) Priority: 17.07.92 JP 212288/92

Date of publication of application : 26.01.94 Bulletin 94/04

(84) Designated Contracting States : DE FR GB IT

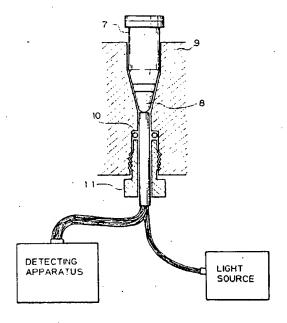
7) Applicant: Tosoh Corporation 4560, Kaisei-cho Shinnanyo-shi, Yamaguchi-ken (JP) 72 Inventor: Mitoma, Yasutami 4-26-5-106, Shonandai Fujisawa-shi, Kanagawa-ken (JP)

(74) Representative: Kearney, Kevin David Nicholas et al KILBURN & STRODE 30 John Street London, WC1N 2DD (GB)

(54) Fluorescence detecting apparatus.

A fluorescence detecting apparatus which allows highly precise measurement of fluorescence even with minute sample amounts, which has a strong responsiveness to temperature variations, which allows simultaneous measurement of a plurality of samples, and wherein the light source and the container holder, and the container holder and the fluorescence detector, are each optically connected by optical fibers; and the optical fibers are connected to the container holder in such a manner that the sample in the container is excited for fluorescence from below the sample container held by the container holder, and that they may receive the fluorescent light which is emitted by the sample from below the sample container.

Fig. 2



The present invention relates to a fluorescence detecting apparatus, and more specifically, it relates to a fluorescence detecting apparatus for measuring variations in the fluorescent properties of substances or their interacted complexes in reactions which occur between at least two types of substances capable of interaction between each other, for example, reactions involving nucleic acids and intercalatory fluorescent pigments, lipid bilayers and hydrophobic fluorescent probes, proteins and fluorescent pigments, organic polymers and fluorescent pigments, etc.

In reactions which occur between at least two types of substances capable of interaction between each other, for example, reactions involving nucleic acids and intercalatory fluorescent pigments, lipid bilayers and hydrophobic fluorescent probes, proteins and fluorescent pigments, organic polymers and fluorescent pigments, etc., the fluorescent properties of these substances or their interacted complexes vary depending upon their state. Thus, if the variation in the fluorescent properties thereof is measured, it is possible to know the state of interreaction of the above substances, the amount of complex formed, etc.

In polymerase chain reactions (PCRs) which are conducted in the copresence of an intercalatory fluorescent pigment (for example Japanese Patent Application Hei 3-313616) etc., it is possible to know the state of nucleic acid amplification (success of the PCR) by measuring the fluorescent properties at a desired point in each cycle of the PCR, and usually at a point during repetition of separation of the double-stranded nucleic acid into single strands and hybridization between single strands, by variation of the temperature. In this procedure, a device is required which can vary the temperature according to a preset program and measure the change in the fluorescent properties of a sample in a sample container. Such a device in current use is a spectrophotometer containing a temperature-controlling cell holder.

15

25

30

35

40

45

50

Measurement of fluorescent property variation according to the prior art using a spectrophotometer employs a system in which exciting light is directed towards a cell using an optical system of lenses, mirrors, etc., and fluorescence is directed towards a fluorescence detecting apparatus in the same manner, and therefore it has been difficult to effect multichannel measurement (simultaneous measurement of multiple samples). Further-more, the responsiveness of the sample temperature to the temperature variation is lessened by the amount of liquid in the sample, and it is difficult to measure the rapid interactions between compounds which accompany phase transitions.

Various attempts have been made at improving temperature responsiveness, but in methods where the volume of the sample container is reduced, etc. it becomes necessary to select, i.e. narrow down, e.g. by using an iris, the beam of the exciting light onto the sample, thus causing new problems such as a reduction in the quantity of light and a resulting decrease in the precision of measurement. An attempt at improvement has been made by using an intense light source to compensate for the reduction in the quantity of light, but in this case the problem of heat waste from the device and the influence of heat on the fluorescence detecting apparatus are considerable, and thus it becomes difficult to obtain highly precise and repeatable measurement results.

Very recently, devices which employ optical fibers in an optical system (Biotechnology, Vol. 10, p.413, 1992) have become known, but in these devices the sample is excited from the top of the sample container, and the fluorescence from the sample is also received from the top, and therefore if the amount of the sample is small, there is a Loss of exciting light and fluorescence from the sample in the air layer between the sample and the tip of the optical fibers. The process is thus complicated by the necessity of replacing the sample container from time to time, depending on the volume of the sample, in order to overcome this problem, and therefore improvement thereof is in order.

Fig. 1 is a drawing showing the construction of an apparatus according to the present invention;

Fig. 2 is a drawing showing the periphery of the container holder of the apparatus in Fig. 1;

Fig. 3 is a graph showing the results of Example 1, with the fluorescence intensity during the PCR cycle for various concentrations of λDNA plotted on the vertical axis, and the number of PCR cycles plotted on the horizontal axis. In the figure, "a" represents a case where the amount of DNA is 2.5 ng, "b" a case where it is 0.25 ng, and "c" a case where it is 0.025 ng.

Fig. 4 is a graph showing the results of Example 2, with the fluorescence intensity during a PCR conducted in the presence of an intercalatory fluorescent pigment plotted on the vertical axis, indicating temperatures of the sample at the time of measurement of the fluorescence intensity on the right-hand side, and the time plotted on the horizontal axis.

We, the present inventors, as a result of intensive research carried out in order to overcome the above mentioned problems, have succeeded in completing the present invention. In other words, the present invention relates to a fluorescence detecting apparatus which comprises a sample container which holds a sample, a container holder which holds the sample container and is capable of varying the temperature of the sample in the sample container which it holds, a fluorescence detector for measuring the fluorescence from the sample, and a light source which emits exciting light to excite the sample for fluorescence, characterized in that the light source and the container holder, and the container holder and the fluorescence detector, are each

the fluorescent light from the sample on the exterior, and the ends of the optical fibers for directing the exciting light in the center, since this allows for reception of weaker fluorescence. As mentioned previously, if the two systems are provided for by a single optical fiber line, then the end of the fiber line may be simply connected to the container holder.

The source of the exciting light is normally selected to maximize the light intensity, but in cases where there exists the possibility of deterioration of the sample due to the radiation of the exciting light, a light-interrupting shutter may be provided to prevent the exciting light from radiating onto the sample, and the shutter may be opened and closed only at the time of measurement of the fluorescence to avoid constant bombardment of the exciting light onto the sample. The light-interrupting shutter is provided in the optical path running from the light source to the container holder, but the simplest construction is one in which a movable shutter is constructed between the ends of the optical fibers and the light source, and the shutter is mechanically or electrically moved to open and close in synchronization with the timing of measurement of the fluorescence. This timing may be, for example, the time at which the temperature of the container holder falls within a desired range (i.e., the time at which the sample in the container is thought to be in the desired range). Such a procedure is particularly effective in cases where continuous (intermittent) detection of fluorescence from two identical samples is made, such as in the fluorescence measurement in PCRs, as disclosed in Japanese Patent Application Hei 3-313616.

The variation of the temperature of the container holder for varying the sample temperature may be effected within a desired range, and this may be stored in a program, and a controller for controlling the temperature of the container holder may be attached to the apparatus according to the present invention. The controller may include a sensor for sensing the temperature of the container holder and means for storing the above mentioned program, and may compare the signal from the above mentioned sensor and the contents of the program, outputting an indication for heating or cooling of the container holder, and this purpose may be achieved by using a microcomputer or the like. If the controller functions to control the detector, light source and shutter, etc. of the present invention, then it is possible to achieve an automatic fluorescence detecting apparatus which varies the temperature of a sample when the sample container is placed in the container holder, and which can measure fluorescence by opening and closing the shutter in accordance with the timing.

20

40

45

The apparatus according to the present invention is suitable for use in PCRs. In this case, it is particularly preferable that the above mentioned sample container be sealed. The PCR is a reaction which amplifies the nucleic acid in a sample, and for the purpose of preventing secondary contamination a fluorescence detecting apparatus according to the present invention which employs a sealed sample container is most effective. This is because, by using a sealed container, it is possible to keep to a minimum the contamination of nucleic acids by other samples, which is a cause of false positivity in PCRs.

In most PCRs, completion is not after a single procedure. This is because the separation of double-stranded nucleic acids into single-stranded nucleic acids by varying the temperature of the sample, annealing of a primer to each of the single-stranded nucleic acids, and the extension reaction of nucleic acids originating from the primer (which results in the appearance of double-stranded nucleic acids) are conducted as one cycle, and the cycle is usually repeated. Thus, an apparatus according to the present invention which is suitable for PCRs, is preferably provided with the above mentioned shutter.

Because the above mentioned cycle is repeated, the temperature variation of the container holder is actually the variation between the temperature at which the nucleic acids can exist in double-stranded form (if the object nucleic acid is double-stranded, then this temperature range is applied at the start of the above mentioned cycle, at the time of extension of the primer, and appearance of the double-stranded nucleic acid resulting from the above mentioned extension) and the temperature at which they can only exist in single-stranded form (this temperature range is applied for division of the double-stranded nucleic acid into single strands).

With reactions involving nucleic acids and intercalatory fluorescent pigments as a case in point, here the fluorescent properties of the fluorescent pigment themselves vary in a temperature-dependent manner, and thus the timing of variation of the temperature of a sample in a PCR and opening and shutting of the above mentioned shutter is preferably such that the measurement is made at a stage wherein a fixed temperature has been reached after completion of a series of temperature variations, rather than during variation of the temperature. Furthermore, in this reaction, since a fluorescent pigment is taken up in the nucleic acid extension reaction which originates with a primer, and the fluorescent properties thereof vary, the reaction may be monitored to determine whether or not the PCR is successful (Japanese Patent Application Hei 3-313616); however, here as well the temperature range may be one in which the nucleic acids can exist in double-stranded form, and a timing which maintains a fixed temperature in the interior of the sample container is preferable. In concrete teams, this is at the moment of completion of the above mentioned cycle.

An explanation of the present invention will now be given with reference to the drawings. Fig. 1 is a drawing

5

10

20

35

the fluorescent light from the sample on the exterior, and the ends of the optical fibers for directing the exciting light in the center, since this allows for reception of weaker fluorescence. As mentioned previously, if the two systems are provided for by a single optical fiber line, then the end of the fiber line may be simply connected to the container holder.

The source of the exciting light is normally selected to maximize the light intensity, but in cases where there exists the possibility of deterioration of the sample due to the radiation of the exciting light, a light-interrupting shutter may be provided to prevent the exciting light from radiating onto the sample, and the shutter may be opened and closed only at the time of measurement of the fluorescence to avoid constant bombardment of the exciting light onto the sample. The light-interrupting shutter is provided in the optical path running from the light source to the container holder, but the simplest construction is one in which a movable shutter is constructed between the ends of the optical fibers and the light source, and the shutter is mechanically or electrically moved to open and close in synchronization with the timing of measurement of the fluorescence. This timing may be, for example, the time at which the temperature of the container holder falls within a desired range (i.e., the time at which the sample in the container is thought to be in the desired range). Such a procedure is particularly effective in cases where continuous (intermittent) detection of fluorescence from two identical samples is made, such as in the fluorescence measurement in PCRs, as disclosed in Japanese Patent Application Hei 3-313616.

The variation of the temperature of the container holder for varying the sample temperature may be effected within a desired range, and this may be stored in a program, and a controller for controlling the temperature of the container holder may be attached to the apparatus according to the present invention. The controller may include a sensor for sensing the temperature of the container holder and means for storing the above mentioned program, and may compare the signal from the above mentioned sensor and the contents of the program, outputting an indication for heating or cooling of the container holder, and this purpose may be achieved by using a microcomputer or the like. If the controller functions to control the detector, light source and shutter, etc. of the present invention, then it is possible to achieve an automatic fluorescence detecting apparatus which varies the temperature of a sample when the sample container is placed in the container holder, and which can measure fluorescence by opening and closing the shutter in accordance with the timing.

The apparatus according to the present invention is suitable for use in PCRs. In this case, it is particularly preferable that the above mentioned sample container be sealed. The PCR is a reaction which amplifies the nucleic acid in a sample, and for the purpose of preventing secondary contamination a fluorescence detecting apparatus according to the present invention which employs a sealed sample container is most effective. This is because, by using a sealed container, it is possible to keep to a minimum the contamination of nucleic acids by other samples, which is a cause of false positivity in PCRs.

In most PCRs, completion is not after a single procedure. This is because the separation of double-stranded nucleic acids into single-stranded nucleic acids by varying the temperature of the sample, annealing of a primer to each of the single-stranded nucleic acids, and the extension reaction of nucleic acids originating from the primer (which results in the appearance of double-stranded nucleic acids) are conducted as one cycle, and the cycle is usually repeated. Thus, an apparatus according to the present invention which is suitable for PCRs, is preferably provided with the above mentioned shutter.

Because the above mentioned cycle is repeated, the temperature variation of the container holder is actually the variation between the temperature at which the nucleic acids can exist in double-stranded form (if the object nucleic acid is double-stranded, then this temperature range is applied at the start of the above mentioned cycle, at the time of extension of the primer, and appearance of the double-stranded nucleic acid resulting from the above mentioned extension) and the temperature at which they can only exist in single-stranded form (this temperature range is applied for division of the double-stranded nucleic acid into single strands).

With reactions involving nucleic acids and intercalatory fluorescent pigments as a case in point, here the fluorescent properties of the fluorescent pigment themselves vary in a temperature-dependent manner, and thus the timing of variation of the temperature of a sample in a PCR and opening and shutting of the above mentioned shutter is preferably such that the measurement is made at a stage wherein a fixed temperature has been reached after completion of a series of temperature variations, rather than during variation of the temperature. Furthermore, in this reaction, since a fluorescent pigment is taken up in the nucleic acid extension reaction which originates with a primer, and the fluorescent properties thereof vary, the reaction may be monitored to determine whether or not the PCR is successful (Japanese Patent Application Hei 3-313616); however, here as well the temperature range may be one in which the nucleic acids can exist in double-stranded form, and a timing which maintains a fixed temperature in the interior of the sample container is preferable. In concrete terms, this is at the moment of completion of the above mentioned cycle.

An explanation of the present invention will now be given with reference to the drawings. Fig. 1 is a drawing

showing the entire body of an apparatus according to the present invention. 1 represents a container holder for holding a sample container, provided with a plurality of concave sections for holding containers to allow measurement of a plurality of samples, and optical fibers 5 on each concave section. 2 represents a fluorescence detector, 3 a light source, 4 a shutter, 5 optical fibers, and 6 a controller. Fig. 1 shows a case wherein the end surfaces of the optical fibers for irradiating exciting light and the end surfaces of the optical fibers for receiving fluorescent light are arranged coaxially. Also, in this embodiment the controller used is a microcomputer, and it controls the container holder, the fluorescence detector, the light source and the shutter.

The temperature of the container holder is varied according to the temperature variation program stored in the controller, in order to vary the temperature of the sample in the sample container (not shown). The light source is on during this time, or is controlled to be on in synchronization with the timing of measurement of the fluorescence. The controller opens and closes the shutter according to a prescribed timing, following the signal from a temperature sensor (not shown), thus exciting the sample and allowing the fluorescence to be measured by the fluorescence detector. In this apparatus, the results of measurement by the fluorescence detector may be read by the controller and later displayed.

Fig. 2 is a drawing showing the periphery of the container holder of the apparatus explained in Fig. 1. 7 represents a sealed sample container, 8 a sample in the container, 9 an aluminum container holder which holds the sample container (only one of the plurality of concave sections is shown), 10 an O-ring and 11 a screw which connects the optical fibers to the container holder. In this embodiment, 48 fibers are bunched together and arranged around the perimeter, with 12 for irradiation of the exciting light at the centre of the bundle and the remaining 36 for receiving the fluorescent light arranged around the outside of the bundle.

#### Examples

5

15

25

30

40

45

50

Examples will now be provided to illustrate the apparatus according to the present invention and measurement of fluorescence using it, without limiting the present invention thereto.

#### Example 1

Using a commercially available DNA synthesis kit (GeneAmp, trade name, product of Takara Brewing Co.) and the apparatus shown in Figs. 1 and 2, amplification of nucleic acids was effected using the PCR (1 cycle = denaturation at 94°C for 1 minute, annealing at 55°C for 1 minute, extension at 72°C for 1 minute (using Taq polymerase), and 30 cycles were repeated in the presence of an intercalatory fluorescent pigment (Pigment 33258, product of Hoechst AG). The reaction solution for the PCR was prepared using a calibrating  $\lambda$ -DNA and 1 set of primer included in the kit, and following the directions for the kit. The fluorescent pigment was added to a concentration of 1  $\mu$ g/ml. The composition of the reaction solution is listed below.

55

	$\lambda$ -DNA (	1 µg. 0.1 µg or 0.	01 µg/ml)	10 µ1	
	10 × re	action buffer solu	ition	10 µl	
	(Sodium chloride			0.5 M)	
5	. (Т	ris-HCl buffer sol	Lution (pH 8.0)	0.1 M)	
	( M	agnesium chloride		15 mM)	
	( G	elatin (weight/vol	Lume)	0.01%)	
10	Intercalatory (final conce	fluorescent pigmer ntration)	nt	l ug/ml	
15	Primer 1 and P	rimer 2	(20 µM)	1 µ1	
	(Primer 1:	$\lambda$ -DNA, corresponding antisense strand $\Omega$	ing to 7131-715 GATGAGTTCGTGTCC	5, GTACAACTGG)	
20	(Primer 2:	$\lambda$ -DNA, corresponds sense strand GCTTA	ing to 7607-763 ATCGAAATCAGCCAC	0, AGCGCC)	
	dNTP mixture	· !	(2.5 mM ea)	8 µl	
	Water			59.5 µ1	
25	Taq polymera	ise	(5 units/µl)	0.5 µl	

In the apparatus shown in Figs. 1 and 2, optical fibers (Superesca, product of Mitsubishi Rayon), a controller (DCP200, product of Yamatake Honeywell) and a light source (50W xenon lamp; light power source: Model C-2576, both products of Hamamatsu Photonics) were used, and the shutter and fluorescence detector were manufactured in-house.

For the three different samples prepared in the above manner having different initial  $\lambda$ -DNA concentrations, 25  $\mu$ l of each reaction solution was sampled in a microcentrifuge tube (product of Biobic), mineral oil was superpositioned on the surface thereof to prevent evaporation of the sample, which was carried on an aluminum block, and the PCR was carried out while raising and lowering the temperature. Exciting light from the xenon lamp light source and passing through a U350 color glass filter was focussed with a lens, directed towards the optical fibers, and irradiated from the bottom of the sample container to the sample. The fluorescence from the sample was received by the optical fibers at the bottom of the reaction container, and the light which passed through a 400 - 450 nm interference filter was measured with the in-house manufactured fluorescence detector which contained a photodiode. The measurement of the fluorescence was made during the final second of the extension reaction of the above mentioned reaction cycle.

The resulting measurement of the variation in fluorescence during the PCR was as shown in Fig. 3. In Fig. 3, the fluorescence intensity during the PCR cycle for each concentration of  $\lambda$ -DNA is plotted on the vertical axis, and the number of cycles is plotted on the horizontal axis, and it is clear from this graph that the number of cycles required for an increase in the fluorescence intensity depended on the concentration of the sample; that is, 3 curves were obtained with different increase points depending on the initial amount of DNA in the sample.

#### Example 2

35

40

45

Using the amplified sample from Example 1, continuous measurement was made of the fluorescence intensity at temperatures (94, 72, 55°C) predetermined for the double-stranded nucleic acid fragments (approximately 500 base pairs) and during a state of continuous variation of the temperature to each one, in the presence of an intercalatory fluorescent pigment.

The resulting measurement of the variation in fluorescence was as shown in Fig. 4. In Fig. 4, the time is plotted on the horizontal axis, and the fluorescence intensity on the vertical axis.

#### Example 3

5

10

20

35

40

55

Of  $\lambda$ -DNA which had been digested with EcoT14I, 1  $\mu$ g was suspended in 25  $\mu$ l of a buffer solution (50 mM of NaCl, 1  $\mu$ g/ml of intercalatory fluorescent pigment (33258, product of Hoechst AG), 10 mM of Tris-HCl, pH 8.5), the suspension was placed in a 25  $\mu$ l microcentrifuge tube (product of Biobic), and the fluorescence intensity was measured using an apparatus similar to the one in Example 2, except that coaxial fibers (E32-CC200, product of Omron) were used. For comparison, the above mentioned optical fibers were affixed to the upper opening of the microcentrifuge tube for measurement in the same manner. For this purpose, the end surfaces of the optical fibers positioned at the liquid surface of the sample in the centrifuge tube and at the above mentioned opening were separated by a gap of 23 mm.

The above mentioned optical fibers consisted of one 1 mm diameter fiber in the center, and 16 lines of 0.25 mm fibers around the periphery, with the center fiber used for irradiating exciting light, and the peripheral fibers used for receiving fluorescent light. In this embodiment, a 50 W xenon lamp is used as the light source, and measurement was made of the exciting light which was transmitted through a U-350 filter and of the fluorescent light which was transmitted through an interference filter (400 - 450 nm).

As a result, a fluorescence intensity of 265 mV was obtained with the apparatus according to the present invention, whereas the fluorescence intensity from the same sample was measured to be 20 mV, or only about 1/13 in comparison, when the optical fibers were attached to the upper opening of the centrifuge tube.

According to the present invention, since the light source, the container holder (i.e., the sample) and the fluorescence detector are all connected by the optical fibers, it is possible to arrange all of these separately so that the heat-emitting light source and the container holder do not affect the fluorescence detector. Furthermore, multichannel measurement, or direction of exciting light from a single light source to a plurality of samples, and direction of fluorescence from a plurality of samples to a detector, may be easily realized. If highly photoconductive fibers are used, then the loss of the exciting light and fluorescent light may be prevented and the light beam may be selected, and therefore it is possible to make high precision measurements even in the case of, for example, minute sample amounts, etc.

According to the present invention, exciting light is irradiated onto the sample container preferably the bottom of a sample container, and fluorescent light is received from the sample container preferably at the bottom of the sample, and therefore the air layer between the sample and the detector may be kept to a minimum. As a result, highly precise measurements are possible particularly in cases where fluorescence is measured from minute sample amounts.

The significance of using minute sample amounts is that the responsiveness of samples to temperature variation may be improved, and therefore that in reactions involving substances which undergo temperature-dependent phase transitions and have been difficult to measure according to the prior art, for example, reactions involving intercalatory fluorescent pigments and nucleic acids, reactions involving lipid bilayers and hydrophobic fluorescent probes, reactions involving proteins and fluorescent pigments, and reactions involving organic polymers and fluorescent pigments, the measurement of rapid interactions between compounds which result from phase interactions due to temperature variations may be made by measuring the variation in the fluorescent properties thereof.

Particularly, if an apparatus according to the present invention is used in a PCR in which an intercalatory pigment is added, and the fluorescence intensity is measured according to a desired timing during each cycle, or the variation in the fluorescence is measured throughout each cycle, then it is possible to know the progress of nucleic acid amplification, i.e., the success of the PCR, from these measurements. Therefore, according to the present invention it is also easily possible to know the success of PCRs, which has conventionally been determined by electrophoresis, etc. This means that waste may be reduced in cases where the initial concentration of the object nucleic acid is high and the PCR continues unnoticed even after saturation of the amplification reaction, resulting in the amplification of non-specific nucleic acids, and further means that the subsequent process of identification of the nucleic acids may be facilitated. If the fluorescent property at an appropriate wavelength for the PCR is measured and a sealed sample container is used, then there is no need to remove the sample from the container for measurement, and this is very effective in avoiding false positivity, particularly in the case of samples with a high probability of secondary contamination, etc.

It is particularly advantageous to supply the exciting radiation and to collect the fluorescent radiation from the bottom of the container when the container has a narrowed or pointed bottom because the sample will be collected together at the bottom of such a contain r but a given volume will extend further up the tube thus making removal of samples easier.

It has been mentioned above that the axes of the ends of the exciting light radiation fibres and the fluorescent light receiving fibres may converge. This convergence may be such that the axes intercept within the sample container and preferably in that region the reof where the sample will be located in use, e.g. near to

the bottom, and with containers having narrowed or pointed ends preferably within such narrowed or pointed region.

In its broadest aspects the invention provides a method of measuring fluorescence from a sample which fluoresces or can be excited to fluoresce which comprises locating the sample in a container which is transparent to the fluorescent radiation or has a region which is transparent, locating one end of a fibre optic cable against the outside of the container or close thereto (e.g. less than 1 mm away from the surface), at the said transparent region and conveying fluorescent radiation along the fibre optic cable to a detector for the fluorescent radiation.

The transparent region is preferably at the base of the container which preferably is curved. The container conveniently is a tube. The container is preferably sealable and is preferably sealed during the detection period.

The end of the fibre optic cable is preferably in contact with the outside of the base of the tube shaped container.

The method preferably also includes leading radiation, effective to excite the sample so that it fluoresces, to a region transparent to the excitation radiation by a fibre optic cable, one end of which is located against the outside of the container, or close thereto (e.g. less than 1 mm away from the surface), at the said transparent region.

Preferably the container is a tube which is transparent to both the fluorescent radiation and the excitation radiation.

Preferably the region at which the fluorescent radiation is detected and the region at which the excitation radiation is imparted are close to each other. For example first fibre optic means which may be a bundle of cables forming a central array can be arranged centrally and second fibre optic means which may also be a plurality of cables can be arranged as an array, e.g. a ring, around the central array. Either the first or second means can be used for the fluorescent radiation. Preferably the surrounding ring, the second means, is used for gathering and receiving the fluorescent radiation.

The method also extends to a procedure in which the temperature of the sample is varied in a controlled way and fluorescence measurements are taken at different temperatures. The method also extends to a procedure in which the excitation radiation is applied only for selected periods of time and not continuously.

The invention in a first apparatus aspect also extends to apparatus for detecting fluorescence from a sample which fluoresces or can be excited to fluoresce which comprises a container for a sample, the container preferably having sealing means, the container being transparent to the fluorescent radiation or having a region which is transparent, means for holding the container, first fibre optic means, fluorescence detector means, means for locating one end of the fibre optic means against the outside of the container or close thereto, e.g. less than 1 mm away from the surface, at the said transparent region, and means for connecting the other end of the first fibre optic means optically, and preferably physically as well, to the fluorescence detector means.

The means for holding the container are preferably provided with temperature controlling means whereby the sample in the container can be heated or cooled.

The means for holding the container may be a block, e.g. able to act as a heat sink, e.g. of metal, to facilitate temperature control. The block may contain a recess into which the sample container is adapted to fit. The block may also afford the means by which the end of the first fibre optic means can be located against th sample container.

The invention in a second apparatus aspect also extends to apparatus for detecting fluorescence from a sample which fluoresces or can be excited to fluoresce which comprises a container for a sample, the container being transparent or having a region which is transparent to radiation effective to excite the sample so that it fluoresces, means for holding the container, second fibre optic means, means for generating the excitation radiation, means for locating one end of the fibre optic means against the outside of the container or close thereto, e.g. less than 1 mm away from the surface, at the said transparent region, and means for connecting the other end of the second fibre optic means optically, and preferably physically as well, to the means for generating the excitation radiation.

The first and second apparatus aspects can be used together and most preferably are so used.

The first end of the first and second fibre optic means may be located next to each other at the sample container for example one may be arranged as a ring or annulus around the other. The means for locating the first end of the first and the second fibre optic means can then be a single structure.

#### Claims

5

10

15

20

25

30

35

40

45

50

55

1. A method of measuring fluorescence from a sample which fluor sces or can be excited to fluoresce which

5

20

25

50

55

comprises locating the sample in a container which is transparent to the fluorescent radiation or has a region which is transparent, locating a first end of first fibre optic means against the outside of the container or close thereto (e.g. less than 1 mm away from the surface), at the said transparent region and conveying fluorescent radiation along the fibre optic means to a detector for the fluorescent radiation.

- 2. A method as claimed in Claim 1 which includes leading radiation, effective to excite the sample so that it fluoresces, to a region transparent to the excitation radiation by fibre optic means, a first end of which is located against the outside of the container, or close thereto (e.g. less than 1 mm away from the surface), at the said transparent region.
- 3. Apparatus for detecting fluorescence from a sample which fluoresces or can be excited to fluoresce which comprises a container for a sample, the container being transparent to the fluorescent radiation or having a region which is transparent, means for holding the container, first fibre optic means, fluorescence detector means, means for locating a first end of the first fibre optic means against the outside of the container or close thereto, e.g. less than 1 mm away from the surface, at the said transparent region, and means for connecting the other end of the first fibre optic means optically, and preferably physically as well, to the fluorescence detector means.
  - Apparatus as claimed in Claim 3 in which the means for holding the container are provided with temperature controlling means.
    - 5. Apparatus for detecting fluorescence from a sample which fluoresces or can be excited to fluoresce which comprises a container for a sample, the container being transparent (or having a region which is transparent) to radiation effective to excite the sample so that it fluoresces, means for holding the container, second fibre optic means, means for generating the excitation radiation, means for locating a first end of the second fibre optic means against the outside of the container or close thereto, e.g. less than 1 mm away from the surface, at the said transparent region, and means for connecting the other end of the second fibre optic means optically, and preferably physically as well, to the means for generating the excitation radiation.
- Apparatus for detecting fluorescence from a sample which fluoresces or can be excited to fluoresce which comprises a container for a sample, the container being transparent, or having a region which is trans-30 parent, to the fluorescent radiation, the container being transparent, or having a region which is transparent, to radiation effective to excite the sample so that it fluoresces, means for holding the container, second fibre optic means, means for generating the excitation radiation, means for locating one end of the second fibre optic means against the outside of the container or close thereto, e.g. less than 1 mm away 35 from the surface, at the said region transparent to the excitation radiation, and means for connecting the other end of the second fibre optic means optically, and preferably physically as well, to the means for generating the excitation radiation, and first fibre optic means, fluorescence detector means, means for locating one end of the first fibre optic means against the outside of the container or close thereto, e.g. less than 1 mm away from the surface, at the said region transparent to the fluorescent radiation, and means for connecting the other end of the first fibre optic means optically, and preferably physically as 40 well, to the fluorescence detector means.
- 7. A fluorescence detecting apparatus which comprises a sample container which holds a sample, a container holder which holds the sample container and is capable of varying the temperature of the sample in the sample container which it holds, a fluorescence detector for measuring the fluorescence from the sample, and a light source which emits an exciting light to excite the sample for fluorescence, characterized in that:

the light source and the container holder, and the container holder and the fluorescence detector, are each optically connected by optical fibers; and

said optical fibers are arranged in the container holder in such a manner that the sample in said container excited for fluorescence from below the sample container held by said container holder, and that they may receive the fluorescent light which is emitted by the sample from below the sample container.

8. A fluorescence detecting apparatus according to any one of Claims 3 to 7, which is provided with at least one light-interrupting shutter in the optical path of the exciting light used to excite the sample for fluorescence, which runs from the light source to the container holder.

9.	A fluorescence detecting apparatus according to any one of Claims 3 to 8 wherein the sample container is a sealed container.
10.	A fluorescence detecting apparatus according to Claim 8 or Claim 9, wherein the light-interrupting shutter is opened and closed in synchronization with the change in the temperature of the container holder.

Fig. 1

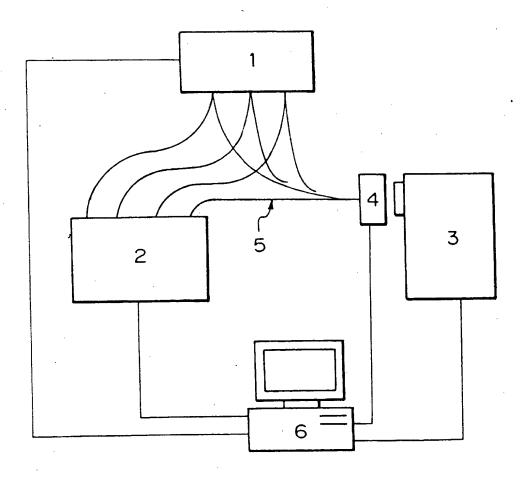


Fig. 2

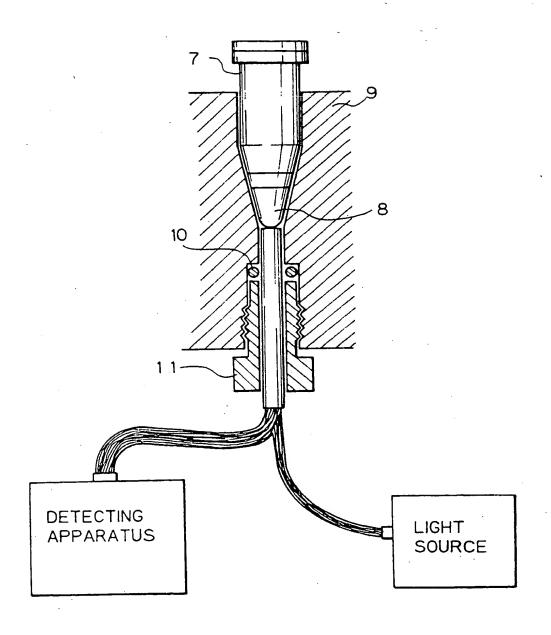
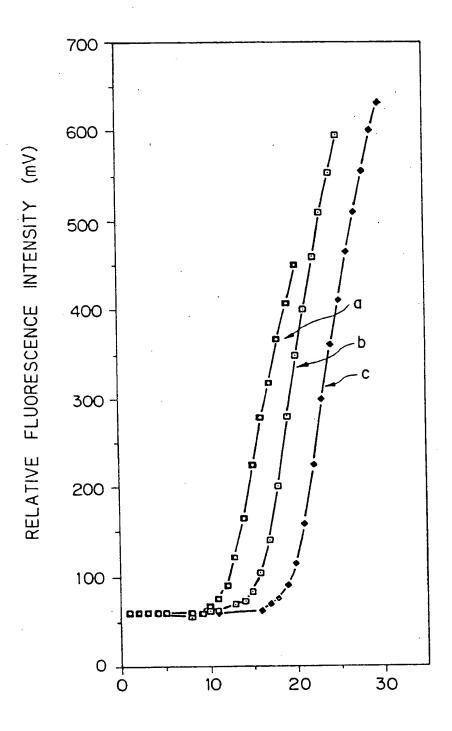
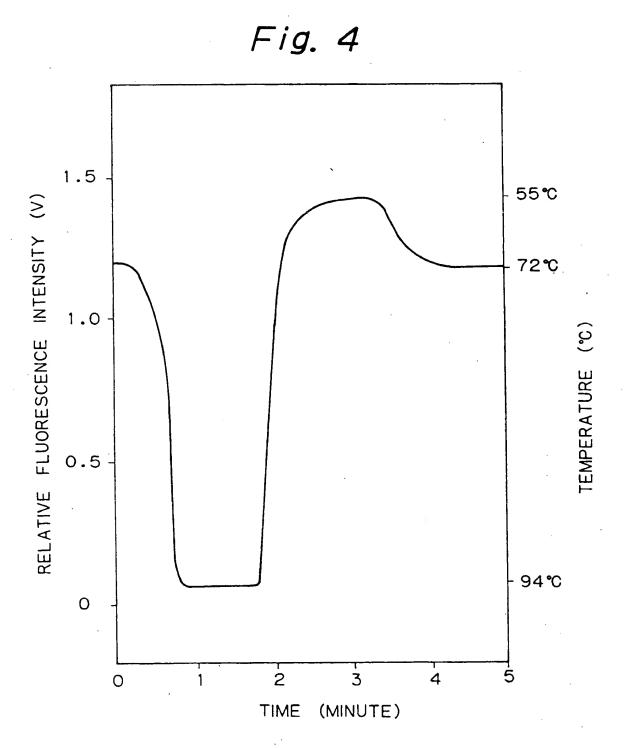


Fig. 3







# **EUROPEAN SEARCH REPORT**

Application Number

EP 93 30 5569 Page 1

	DOCUMENTS CONSIDE	ation, where appropriate,	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5)	
ategory	of relevant passa	ges			
` i	US-A-3 992 631 (HARTE	)	1-3,5,6	G011(21) 04	
	* abstract *	14 24 *			
	* column 4, line 19 -	column 7 line 5:	3,4		
ľ	* column 6, line 38 -	Column 7, Time 3,			
i	figures 1,2 *		ļ	-	
J	EP-A-0 266 881 (ASTRO	MED)	1,2	:	
X	* column 3, line 18 -	· line 43 *		: :	
Υ	* figure 1 *		7	Ţ.	
,	119070 1				
D,Y	BIOTECHNOLOGY	•	3,4,7		
- , -	vol. 10, April 1992,		1		
	names 413 - 417				
	D HUGUCHT FT AL. 'ST	imultaneous			
	amplification and det	tection of specific			
	DNA sequences' * page 415, left colu	mn line 14 - line		,	
	* page 415, left con	Jim, Time 14			
	23; figure 5 *			_	
A	EP-A-0 127 418 (LABS	YSTEMS)	1-3,5-	7	
^	* abstract *			TECHNICAL FIELDS SEARCHED (Int. Cl.5.)	
	* page 4. line 3 - 1	ine 8 *		SEARCHED (IIII OII)	
	* nage 4 line 36 -	page 5, line 1 ^		GOIN	
	* page 5, line 29 -	line 30; figure 2 ~		GOTIV	
			1-7,9	•	
A	WO-A-8 706 716 (BAXT	ing 19 *	1 ,,,,		
	* page 8, line 9 - 1	line 14 *			
1	* page 9, line 12 - * page 13, line 4 -	line 28 *			
,	* page 15, line 18 -	· line 21 *		, ,	
1	* page 18, line 29 -		ļ		
	* figures 1-4,7 *			i •	
			, -		
A	EP-A-0 127 286 (TOSF	HIBA)	1-7	1	
	* nage 1. line 13 -	line 16 "	1		
	* page 5, line 29 -	page b, line 25;			
	figure 4 *				
	ì	-/	}		
		,	Ì		
	·		İ		
	The present search report has b			Examiner	
	Place of search	Date of completion of the se		THOMAS R.M.	
ē i	THE HAGUE	26 OCTOBER 199			
<u>.</u>	CATEGORY OF CITED DOCUME	NTS T: theory o	r principle underlyi	ng the invention	
ntier the fil				ent document, but published on, or ling date	
S X	particularly relevant if combined with an		D: document cited in the application L: document cited for other reasons  4: member of the same patent family, corresponding		
Ξ. A	document of the same category rechnological background				
S 0	non-written disclosure intermediate document	gocume a compa		-	



## **EUROPEAN SEARCH REPORT**

Application Number

EP 93 30 5569 Page 2

	DOCUMENTS CONSIDE			A ACCIDION OF THE	
ategory	Citation of document with indica of relevant passag	ation, where appropriate,	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.5.)	
P, X	EP-A-0 516 274 (KOWA)		5	•	
	<pre>* abstract * * column 4, line 5 -</pre>	line 8 *			
	* column 4. line 28 -	line 31 *		i	
	* column 4. line 54 -	column 5, line 3	*		
	* column 5, line 43 -	line 48 *	3,4,6,7	•	
P,A	* figures 1-3 *		3,4,0,7		
	-				
				· ·	
	•				
	<u> </u>				
			,		
				,	
	i !		1		
				TECHNICAL FIELDS SEARCHED (Int. Cl.5 )	
				SEARCHED (IBC. Cl.5)	
İ					
	:			i	
				1	
	-				
	i				
	1				
	The present search report has been drawn up for all claims				
	Pixe of searth	Date of completion of the 26 OCTOBER 19		THOMAS R.M.	
	THE HAGUE				
X: Y: O: P:	E : earlier pi		patent document, but p	principle underlying the invention tent document, but published on, or	
X	particularly relevant if taken alone particularly relevant if combined with anot	D : doesm	he filing date ent cited in the applicat	ing date	
A :	document of the same category	17. 000.000	L: document cited for other reasons		
Q:	non-written disclosure intermediate document	&:membe docum	& : member of the same patent family, corresponding document		